Bioaerosol Sampling Report

FCC Environment

Bryn Lane IVC (Phase 1)

16th April 2026

Approval Sheet

Customer:	FCC Environment
Site:	Bryn Lane IVC, Wrexham Recycling Park, Bryn Lane, Wrexham LL13 9UT
Project Title:	Bioaerosols Sampling
Project Manager:	A. Aitken
Project Staff:	R. Chau S. Mullen L. Kiernan
Address:	HSL Compliance Enitial Ltd Enterprise Drive Four Ashes Wolverhampton WV10 7DE Tel: 01902 798798 Fax: 01902 798711

Issue	Status	Date	Prepared By	Signature	Date
1	Final	22/04/2026	A. Aitken	A. S. Aitken	08/05/2026
			Approved By	**Signature**	**Date**
			Dr J. Taylor		08/05/2026

Foreword

HSL Compliance Enitial has used its best endeavours, experience and expertise to provide a meaningful, accurate and relevant representation of the works carried out. The works were based on a defined programme and scope of works and terms and conditions agreed with the Client.

HSL Compliance Enitial cannot accept responsibility to any parties whatsoever, following the issue of this report, for any matters arising which may be considered outside the agreed scope of works.

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1.0 Scope of Work

HSL Compliance Enitial was tasked with providing Bioaerosol monitoring for FCC Environment at Bryn Lane IVC (Phase 1) located within Wrexham Recycling Park, Bryn Lane, LL13 9UT.

Due to the potential health risks posed by the facility to nearby receptors, the testing was conducted to assess the amount of airborne bioaerosols being generated by the site that is released into the local environment. The monitoring is to ensure that this exposure is below the industry standard threshold set by the Environmental Agency.

Due to the complex layout and proximity of the site facility to other buildings an agreement has been made between the regulator Natural Resources Wales (NRW) and FCC Environment Ltd. This is an amendment to the standards by the Environment Agency Technical Guidance Note (Monitoring) M9 to produce a modified site strategy.

FCC agree with NRW that the nearest sensitive receptor to the composting operation for sampling will be the Council Yard & Offices, which are approximately 70-80m from the emission stack.

The work was carried out on 16th April 2026. This document is a presentation of the resulting data.

2.0 Background

The bioaerosol monitoring was specifically undertaken to enumerate the quantity of micro-organisms that can be cultured from representative samples of ambient air, collected at pre-determined monitoring points in the locality of the site.

The monitoring was undertaken in accordance with Environment Agency Technical Guidance Note (Monitoring) M9 – Environmental monitoring of bioaerosols at regulated facilities – July 2018.

Pre-prepared agar sample plates for *Aspergillus fumigatus,* mesophilic micro-organisms (bacteria) were directly impacted with ambient air using an Andersen sampler at the four selected sampling locations.

In the UK, there are currently no statutory limits for ambient concentrations of bioaerosols. However, the Environment Agency's former Technical Guidance Note M17 provided guideline levels for assessing acceptable levels of bioaerosol concentrations at sensitive receptors:

- Total bacteria: 1000 cfu/m³
- Aspergillus fumigatus: 500 cfu/m³
- Gram Negative Bacteria: 300 cfu/m^3

Although M17 was withdrawn in October 2024 and replaced by new guidance which does not specify acceptable concentration levels. Therefore, until updated guidance is issued, the M17 guideline values continue to be used.

3.0 Methodology

3.1 Sampling Locations

Sample locations should be determined prior to the commencement of the sampling event where possible. The sampling locations have been set by NRW and FCC and were determined to achieve:

- Upwind sample location 001: One sampling location directly upwind of the centre of the IVC facility at the emissions stack.
- Downwind sample location 002: One location directly downwind of the IVC facility at the emissions stack.
- Sensitive Receptor sample location 003: One location at the nearest sensitive receptor, being the Council Yard and Offices.

3.2 Agar Plates

The types of Agar used were:

Mesophilic micro-organisms: (Total Viable Count [TVC]) – Half Strength Nutrient Agar (also known as 93's – white in colour)

Aspergillus Fumigatus / Total Fungi: (colony-forming unit [cfu]) – Malt Extract Agar (also known as 94's – light yellow in colour)

Gram Negative: MacConkey (MAC) Agar + 0.2 g/L cyclohexamide (also known as VRBGA - pink in colour)

The impacted agar plates were subsequently delivered to a specialist laboratory within 24 hours via a cool box with ice packs where they were cultured and enumerated.

3.3 Equipment

3.3.1 List of Equipment

The equipment used is as follows:

- Continuous operation mobile weather station (wind speed, direction, temperature, humidity) and tripod
- Digital stopwatch

- GPS device
- 4 x single-stage Andersen samplers
- 4 x tripods
- 4 x hemicylindrical baffles
- 4 x stoppers for Andersen samplers
- 4 x fully charged vacuum pumps (individual capacity of at least 35l per min) and connecting tubing
- Rotameter to fit vacuum pump
- Agar plates – variety dependent on required test
- 2 x sealable airtight sterile plastic containers
- Cool box with ice packs for transport
- 70% v/v aqueous solution / wipes of ethanol or industrial methylated spirits cleaning solution or other suitable disinfectants

3.3.2 Cleaning of Equipment

All parts of the sampling equipment that were in contact with the samples, work surfaces or storage containers were cleaned with a 70% aqueous solution or wipes of ethanol or methylated spirits, or other suitable disinfectants prior to use. Equipment was completely dried, through air drying before the sampling exercise was commenced.

On completion of cleaning the Andersen single-stage samplers, a stopper was placed in the cone entrance to stop any contamination occurring before the sampling event.

3.3.3 Setup and Operation

- The pumps were Pre-run for a specified time at the required flow rate of 28.3l/min (±2%). This pump test was undertaken once before the start of the monitoring exercise.
- The Andersen single-stage samplers were mounted on to the tripods securely at a height of 1.5-1.8m above ground and the baffles were fitted on the tripod to form a rear shield when aligned to the desired sampling direction.
- The vacuum pump was connected to the rotameter using an appropriate length and diameter of the tubing. Using the rotameter, the flow rate was adjusted to ensure it is running at a constant flow of 28.3l/min (±2%) for each pump.
- On completion of the pump test, the tubing from the dry gas flow meters was disconnected from the rotameter then connected to the inlet on the corresponding Andersen single-stage sampler.
- Fresh sample plates were installed in turn by removing the lid and being placed within the Andersen single-stage sampler
- The stoppers in the inlet of the Andersen single-stage samplers were removed.

- When the Andersen single-stage samplers were fitted with plates the vacuum pumps were activated and the start time recorded on the stopwatch. All pumps were switched on/off at the same time or at a maximum interval of up to 10% of the total run time.
- When the sample duration was completed, the pumps were stopped, and the finish time recorded.
- The plates were carefully removed ensuring that no contact was made with the exposed agar surface,
- The dish cover was replaced on to the plate and secured with masking tape.
- Each plate was placed into an individual plastic bag and sealed.
- The plates were stored in an upright, protective and cooled container and transported to the laboratory within 24 hours.
- Control blank samples were taken at the site. The steps stated above were adhered to however the sample pump was not switched on. One of each sample media was inserted within the Andersen single-stage samplers, left in for the same period of sampling time, then packaged.
- Field blank samples were taken. The sample media were placed in re-sealable packaging without being opened.

3.3.4 Laboratory

The impacted agar plates were subsequently delivered to a specialist laboratory via a cool box with an ice pack. The analysis took place within 24 hours from sample collection.

The agar samples were then cultured and counted by a specialist laboratory. The count result provides a quantification of the potential health risks posed by the facility to nearby receptors.

4.0 Presentation of Data

The following pages consist of meteorological conditions found on site, a presentation of the estimated concentrations of airborne micro-organisms, a plan displaying sample locations and the laboratory data.

4.1 Field Sampling Report

Site: Bryn Lane (Phase 1) IVC Facility

Site Operator: FCC Environmental

Date: 16th April 2026

Start Time: 09:00
Finish Time: 10:30

Monitoring Technician ID: R. Chau, S. Mullen and L. Kiernan

4.1.1 Meteorological Conditions

Table 1: Weather Conditions

Parameter	Wind Direction	Wind Speed (m/s)	Pressure (mb)	Temperature (°C)	Cloud Cover (0-8)
Start	SW	5.3	1017	11	2
Finish	SW	5.3	1017	11	2

4.1.2 Site Description

FCC Bryn Lane (Phase 1) IVC facility is part of Wrexham Recycling Park in Wrexham Industrial Estate located on Bryn Lane, Wrexham in North Wales.

The Recycling Park also includes a Materials Recycling facility (MRF), (Phase 2) and a Household Waste Recycling Centre (HWRC) facility operated by FCC Environment. Wrexham Industrial Estate includes a nearby large food production facility and business park.

The IVC facility was fully operational during the sampling period and operating under normal conditions. The nearest sensitive receptor is the council yard and offices located Southwest of the IVC.

4.2 Sampling Locations & Field Comments

Samples were taken at an upwind, downwind location and at a sensitive receptor (council yard and offices) (see enclosed annotated image Appendix 'A') using Andersen samplers. Ambient air samples were collected by the technique of direct impaction, where a known quantity of air was directed onto an agar plate. The agar samples were then cultured and counted by a specialist laboratory. The count result provides a quantification of the potential health risks posed by the facility to nearby receptors.

Table 2: Sampling Locations

Location	Description
Upwind location 001	Grass verge between bay 6 and 7
Downwind location 002	Grass path behind IVC
Sensitive Receptor location 003	Grass verge close to fence line

An aerial plan, as well as photographs of the locations are presented in *Appendix A* and *Appendix B* respectively

4.3 Deviations from Methods

The locations were assessed to take into account the effect of features and obstructions which may impact the effectiveness of the monitoring. The sampling locations were restricted to where safe physical access was possible along with being legally accessible. Any deviations to the locations, as well as other potential impacts to the assessment are presented in *Table 3* below.

Table 3: Deviations

Location	Deviation	Reason
Upwind Location	Limited distance from the facility and sample location.	No access beyond site boundary and site infrastructure.
Downwind Location	Limited distance from the facility and sample location.	No access beyond site boundary and site infrastructure.

4.4 Field Sampling Record

Table 4: Field Sampling Results

Ambient Sampling: Estimated concentration of bioaerosols										
Site: Bryn Lane IVC (Phase 1) Sampling Date: 16/04/2026 Estimated Mass of Materials: 0 tonne green waste, 120 tonne MBT Activities affecting the concentration of Bioaerosols: IVC activities, biofilter stack				Site Operator: FCC Environment Monitoring Contractor: HSL Compliance Enitial Type of Materials Processed on Site: Green Waste and Compost Site Activity: Normal IVC operations						
Location and grid reference	Sample reference number	Distance from centre of active area (m)	Difference in bearing between location of samplers and mean direction wind blows to (°)	Sampling start/end times (HH:MM)	Concentration of bioaerosols (CFU/m^3) Total Count (TVC)	Concentration of bioaerosols (CFU/m^3) Aspergillus Fumigatus (YM)	Concentration of bioaerosols (CFU/m^3) Gram Negative Bacteria (GN)	Median of samples		
								TVC (CFU/m^3)	YM (CFU/m^3)	GN (CFU/m^3)
Upwind (1) SJ 38616 49924	UW001BIOAEM Upwind of IVC emissions stack	42m	35°	09:30-09:35	57	-	-	28	7	0
				09:36-09:41	-	7	-			
				09:42-09:47	-	-	0			
				09:48-09:53	0	-	-			
				09:54-09:59	-	14	-			
				10:00-10:05	-	-	0			
				10:06-10:11	28	-	-			
				10:12-10:17	-	0	-			
				10:18-10:23	-	-	0			

Location and grid reference	Sample reference number	Distance from centre of active area emission stack (m)	Difference in bearing between location of samplers and mean direction wind blows to (°)	Sampling start/end times (HH:MM)	Concentration of bioaerosols (CFU/m^3) Total Count (TVC)	Concentration of bioaerosols (CFU/m^3) Aspergillus Fumigatus (YM)	Concentration of bioaerosols (CFU/m^3) Gram Negative (GN)	Median of samples		
								TVC (CFU/m^3)	YM (CFU/m^3)	GN (CFU/m^3)
Downwind (2) SJ 38700 49941	DW002BIOAEM Downwind of IVC emissions stack	44m	210°	09:30-09:35	28	-	-	28	28	0
				09:36-09:41	-	28	-			
				09:42-09:47	-	-	0			
				09:48-09:53	7	-	-			
				09:54-09:59	-	7	-			
				10:00-10:05	-	-	7			
				10:06-10:11	35	-	-			
				10:12-10:17	-	28	-			
				10:18-10:23	-	-	0			
Receptor (3) SJ 38661 49910	SR003BIOAEM Receptor location on grassy area south of IVC near boundary to council offices	42m	293°	09:30-09:35	106	-	-	21	0	0
				09:36-09:41	-	0	-			
				09:42-09:47	-	-	14			
				09:48-09:53	21	-	-			
				09:54-09:59	-	0	-			
				10:00-10:05	-	-	0			
				10:06-10:11	21	-	-			
				10:12-10:17	-	0	-			
				10:18-10:23	-	-	0			

Location and grid reference	Sample reference number	Distance from centre of active area (m)	Difference in bearing between location of samplers and mean direction wind blows to (°)	Sampling start/end times (HH:MM)	Concentration of bioaerosols (CFU/m^3) Total Count (TVC)	Concentration of bioaerosols (CFU/m^3) Aspergillus Fumigatus (YM)	Concentration of bioaerosols (CFU/m^3) Gram Negative (GN)	Median of samples		
								TVC (CFU/m^3)	YM (CFU/m^3)	GN (CFU/m^3)
Control	ControlBIOAEM Not removed from package. For QA/QC	N/A	N/A	N/A	0	-	-	N/A	N/A	N/A
					-	0	-			
					-	-	0			
Field	FieldBIOAEM. At same location as Upwind	N/A	N/A	N/A	0	-	-	N/A	N/A	N/A
					-	0	-			
					-	-	0			

4.5 Process Contribution Results

The median is routinely used for statistics and probability theory as the results are less likely to be skewed by extremely high or low values that are not representative of the data set. The median is located by finding the middle value by evenly separating the data set. The median for each location has been calculated and then the highest result for the downwind locations identified. The process contribution has then been calculated by subtracting the upwind median value from the highest downwind median value. The process contribution shows the input the activity of the site has on the ambient bioaerosols concentrations. In the table below the process contribution is shown.

Table 5: Process Contribution Results

Process Contribution Results				
Sample type	Upwind median results	Downwind median highest results	Process Contribution	Sensitive Receptor
Total Bacteria viable Count (TVC)	28	28	**0**	21
Aspergillus fumigatus (YM)	7	28	**21**	0
Gram Negative Bacteria (GN)	0	0	**0**	0

5.0 Summary of Analytical Results

The above reports indicate that at the time of sampling:

1. Mesophilic micro-organisms - Total Viable Count (TVC) shows consistent readings at the downwind locations compared to the upwind location.

2. Aspergillus readings were higher at the downwind locations compared to the upwind location but not detected at the sensitive receptor location.

3. Gram Negative Bacteria (GN) results were not detected at the upwind, downwind or at the sensitive receptor location.

4. Upwind location median results were **below** the industry standard threshold values of 1000cfu/m^3 for Total Bacteria.

5. Downwind location median process contribution results were **below** the industry standard threshold values of 1000cfu/m^3 for Total Bacteria.

6. Sensitive receptor location median process contribution results were **below** the industry standard threshold values of 1000cfu/m^3 for Total Bacteria.

7. Downwind location median process contribution results of 21cfu/m^3 remained **below** the industry standard threshold values of 500cfu/m^3 for Aspergillus Fumigatus.

8. Sensitive receptor location median results remained **below** the industry standard threshold values of 500cfu/m_3 for Aspergillus Fumigatus.

9. Upwind, downwind and sensitive receptor location were **below** the industry standard threshold values of 300cfu/m^3 for Gram Negative bacteria.

10. Control and Field Blanks for QA/QC showed no evidence of contamination.

APPENDIX A

Aerial Plan

Key

1 – Upwind

2 – Downwind

3 – Sensitive Receptor

Blue area – Emission source

Red Box – Site area



FCC Bryn Lane Phase 1 IVC 16/4/2026
Aerial Plan Showing Bioaerosol Sample Locations

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APPENDIX B
Meteorological Conditions

METEOROLOGICAL CONDITIONS

SITE:	Bryn Lane IVC (Phase 1)	SITE OPERATOR:	FCC Environmental
SAMPLING DATE:	16/04/2026	COMMISSIONING LABORATORY:	BioDet
ESTIMATED MASS OF MATERIALS:	Green Waste IVC 0 MBT 120T	TYPE OF MATERIALS PROCESSED ON SITE:	Green waste and compost

Location	Sample Reference Number	Bearing of samplers from boundary of operational area or turning/ screening operation (° from true north) - GPS	Mean direction the wind blows to during the sampling period (° from true north)	Difference in bearing between location of samples from boundary/ source and mean direction wind blows to (°)	Mean wind speed during sampling (m/s)	Arithmetic mean of air temperature (°C)	Arithmetic mean of relative humidity (%)
Upwind	UW001BIOAEM	80 SJ 38616 49924	45	35	5.3	11	76
Downwind	DW002BIOAEM	255 SJ 38700 49941	45	210	5.3	11	76
Sensitive Receptor	SR001BIOAEM	335 SJ 38661 49910	45	293	5.3	11	76

APPENDIX C
Photograph Sheet

Attachment (images)





001 Upwind





002 Downwind





003 Sensitive Receptor

APPENDIX D
Laboratory Certificate

University of Hertfordshire
Hatfield Herts
AL10 9AB

Biodet
Direct line 01707 284545
Laboratory 01707 284522
Fax 01707 285046

Al Aitken
Enitial,
Enterprise Drive,
Four Ashes
Wolverhampton,
WV10 7DE

Our Ref: ENI/26/1184

Date: 1st May 2026

BIOAEROSOL EXPOSURE REPORT: Bryn Lane Phase 1 IVC

Log No. 1184
Sample date. 16th April 2026
Engineer: Al Aitken
Order no.: ENI 108502

Fourteen nutrient agar and fourteen malt extract agar plates were received on 17th April 2026 following TGN M9 bioaerosol sampling using an Andersen sampler.

Environmental monitoring events were monitored during operations:

	Sample Plate no.	Identification	Analysis required	Volume
1	No.1	P1 UW001BIOAEM (93.)	Nutrient agar	141.5
2	No.2	P1 UW001BIOAEM (94.)	Aspergillus fumigatus	141.5
3	No.3	P1 UW001BIOAEM (VR.)	MacConkey Agar	141.5
4	No.4	P1 UW001BIOAEM (93.A)	Nutrient agar	141.5
5	No.5	P1 UW001BIOAEM (94.A)	Aspergillus fumigatus	141.5
6	No.6	P1 UW001BIOAEM (VR.A)	MacConkey Agar	141.5
7	No.7	P1 UW001BIOAEM (93.B)	Nutrient agar	141.5
8	No.8	P1 UW001BIOAEM (94.B)	Aspergillus fumigatus	141.5
9	No.9	P1 UW001BIOAEM (VR.B)	MacConkey Agar	141.5
10	No.10	P1 DW002BIOAEM (93.)	Nutrient agar	141.5
11	No.11	P1 DW002BIOAEM (94.)	Aspergillus fumigatus	141.5
12	No.12	P1 DW002BIOAEM (VR.)	MacConkey Agar	141.5
13	No.13	P1 DW002BIOAEM (93.A)	Nutrient agar	141.5
14	No.14	P1 DW002BIOAEM (94.A)	Aspergillus fumigatus	141.5
15	No.15	P1 DW002BIOAEM (VR.A)	MacConkey Agar	141.5
16	No.16	P1 DW002BIOAEM (93.B)	Nutrient agar	141.5
17	No.17	P1 DW002BIOAEM (94.B)	Aspergillus fumigatus	141.5
18	No.18	P1 DW002BIOAEM (VR.B)	MacConkey Agar	141.5

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19	No.19	P1 DW003BIOAEM (93.)	Nutrient agar	141.5
20	No.20	P1 DW003BIOAEM (94.)	Aspergillus fumigatus	141.5
21	No.21	P1 DW003BIOAEM (VR.)	MacConkey Agar	141.5
22	No.22	P1 DW003BIOAEM (93.A)	Nutrient agar	141.5
23	No.23	P1 DW003BIOAEM (94.A)	Aspergillus fumigatus	141.5
24	No.24	P1 DW003BIOAEM (VR.A)	MacConkey Agar	141.5
25	No.25	P1 DW003BIOAEM (93.B)	Nutrient agar	141.5
26	No.26	P1 DW003BIOAEM (94.B)	Aspergillus fumigatus	141.5
27	No.27	P1 DW003BIOAEM (VR.B)	MacConkey Agar	141.5
28	No.28	P1 CONTROLBIOAEM (93.C)	Nutrient agar	141.5
29	No.29	P1 CONTROLBIOAEM (94.C)	Aspergillus fumigatus	141.5
30	No.30	P1 CONTROLBIOAEM (VR.C)	MacConkey Agar	141.5
31	No.31	P1 FIELDBIOAEM (93.F)	Nutrient agar	141.5
32	No.32	P1 FIELDBIOAEM (94.F)	Aspergillus fumigatus	141.5
33	No.33	P1 FIELDBIOAEM (VR.F)	MacConkey Agar	141.5

Nutrient agar (NA) agar plates were used for total mesophilic bacteria.
Malt extract agar (MEA) agar plates were used for *Aspergillus fumigatus*.
MacConkey agar (MacA) agar plates were used for Gram-negative bacteria.

Samples were incubated for 5 days at 37C (total mesophilic bacteria); for 2 days at 41C (*Aspergillus fumigatus*), and for 2 days at 37C (Gram-negative bacteria).

Results:
Site **Bryn Lane Phase 1 IVC**
Date 16th April 2026
Comments: All culture plates were received in good condition.

Table 1. Microbiological Results:

Sample no.	Sample Head no.	Location	Total Mesophilic Bacteria (cfu per plate)	*Aspergillus Fumigatus* (cfu per plate^3)	Gram-negative bacteria (cfu per plate^3)
1	No.1	P1 UW001BIOAEM (93.)	8		
2	No.2	P1 UW001BIOAEM (94.)		1	
3	No.3	P1 UW001BIOAEM (VR.)			0
4	No.4	P1 UW001BIOAEM (93.A)	0		
5	No.5	P1 UW001BIOAEM (94.A)		2	
6	No.6	P1 UW001BIOAEM (VR.A)			0
7	No.7	P1 UW001BIOAEM (93.B)	4		
8	No.8	P1 UW001BIOAEM (94.B)		0	

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9	No.9	P1 UW001BIOAEM (VR.B)			0
10	No.10	P1 DW002BIOAEM (93.)	4		
11	No.11	P1 DW002BIOAEM (94.)		4	
12	No.12	P1 DW002BIOAEM (VR.)			0
13	No.13	P1 DW002BIOAEM (93.A)	1		
14	No.14	P1 DW002BIOAEM (94.A)		1	
15	No.15	P1 DW002BIOAEM (VR.A)			1
16	No.16	P1 DW002BIOAEM (93.B)	5		
17	No.17	P1 DW002BIOAEM (94.B)		4	
18	No.18	P1 DW002BIOAEM (VR.B)			0
19	No.19	P1 DW003BIOAEM (93.)	15		
20	No.20	P1 DW003BIOAEM (94.)		0	
21	No.21	P1 DW003BIOAEM (VR.)			2
22	No.22	P1 DW003BIOAEM (93.A)	3		
23	No.23	P1 DW003BIOAEM (94.A)		0	
24	No.24	P1 DW003BIOAEM (VR.A)			0
25	No.25	P1 DW003BIOAEM (93.B)	3		
26	No.26	P1 DW003BIOAEM (94.B)		0	
27	No.27	P1 DW003BIOAEM (VR.B)			0
28	No.28	P1 CONTROLBIOAEM (93.C)	0		
29	No.29	P1 CONTROLBIOAEM (94.C)		0	
30	No.30	P1 CONTROLBIOAEM (VR.C)			0
31	No.31	P1 FIELDBIOAEM (93.F)	0		
32	No.32	P1 FIELDBIOAEM (94.F)		0	
33	No.33	P1 FIELDBIOAEM (VR.F)			0

Results are expressed as total micro-organisms per culture plate collected during the exposure time.



1st May 2026

Richard Smith
DIRECTOR OF BIODET